INTRODUCTION TO REVERSE TRANSCRIPTION PCR (RT-PCR)

ABCF 2016 BecA-ILRI Hub, Nairobi 21st September 2016

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Objective of PCR

To provide a solution to one of the most pressing problems facing biology at the time: the replication of DNA.

To eliminate the use of radioisotopes or toxic chemicals.

To amplify a given region of DNA (region of interest).

Polymerase Chain Reaction (PCR) conceptualized in 1983 by American biochemist Dr Kary Banks Mullis

Nobel Prize-winner in chemistry in 1993, for the invention of the PCR.



Theory of PCR

- The Driver of PCR is the **Polymerase** enzyme
- A polymerase will synthesize a complementary sequence of bases to any single strand of DNA providing it has a double stranded starting point
 Polymerase Chain Reaction PCR

- Any gene can be specifically amplified by the polymerase in a mixed DNA sample by adding small pieces of complementary DNA
- These small pieces of DNA are known as **primers** because they prime the DNA sample ready for the polymerase to bind and begin copying the gene of interest.
- During a PCR, changes in temperature are used to control the activity of the polymerase and the binding of primers.

Reverse transcription polymerase chain reaction (RT-PCR)

- 1. RT-PCR refers to PCR that uses product of an Reverse Transcription (RT) reaction as template
- 2. A variant of polymerase chain reaction (PCR)
- 3. A technique commonly used in molecular biology to detect <u>RNA</u> expression
- 4. RT-PCR is often confused with <u>real-time polymerase chain</u> reaction (qPCR)
- 5. RT-PCR is used to **qualitatively** detect gene expression through creation of <u>complementary DNA</u> (**cDNA**) transcripts from RNA
- **6. qPCR** is used to quantitatively measure the amplification of DNA using fluorescent dyes

RT-PCR Principle



Some priming methods for generating first strand cDNA:

Simplicity

- Oligo(dT)-based priming: RNA with a poly(A) tail
- 2. Random hexamer priming: fragmented RNA (<500 b)
- 3. Gene-specific priming: low abundance RNA

RT-PCR is also used in gene cloning, transcriptomics;

One-Step vs Two-Step RT-PCR





One-Step Cycling Conditions



RT-PCR: Contamination gDNA

All RNA Isolation Methods Yield RNA Containing Residual Genomic DNA

- Should avoid detecting DNA contamination;
- Remove DNA:
 - DNase I treatment then phenol:CHCl3 extraction;
 - Lithium chloride (2.5M) precipitation then ethanol wash (but not very efficient for small RNA (<200 nt) and dNTPs; LiCl does not efficiently precipitate DNA, protein or carbohydrate;
 - Poly(A) RNA purification;
- Use primers that span an intro/exon boundary;

Use primers that span an intro/exon boundary

Tp5 (putative translation initiation factor eIF-1A)



155 aa

Principle of PCR 1

- 1. The reaction's temperature is raised to 95°C to denature all double stranded DNA into single strands: **Denaturation**
- 2. The temperature is then lowered to 55-65°C to allow the primers to bind to your gene of interest: **Annealing**
- The optimal temperature for the Taq to operate is 72°C. So the temperature is raised to 72°C: Extension There are now twice as many copies of your gene of interest as when you started
- 4. To do its job the Taq requires a supply of DNA building blocks, i.e., the nucleotides: A, T, C and G

Principle of PCR 2

 The cycle of changing temperatures (95°C, 55°C and 72°C) is then repeated and two copies become four. Another cycle and four become eight, up to 30-35 cycles. After 'n' cycles = 2⁽ⁿ⁺¹⁾ copies



Thermal Cycler

Also known as:

- Thermocycler
- PCR machine
- DNA amplifier



A very early PCR machine Samples moved mechanically







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PCR Cycling



DNA Copy Types:1. Target DNA2. Long-length DNA



Complete PCR

After 30 cycles, from only one molecule: 1,073,741,764 copies of target DNA 60 longer molecules





Analysis of PCR products

After amplifying your gene into many millions of copies, it is possible to run the amplicons on an agarose gel and stain it with a dye to visualize it.



M 1 2 3 4 5 6 7 8 9



Play Principle of PCR Movie



Thank you







